Review

Influence of pre- and peri-natal nutrition on skeletal acquisition and maintenance☆

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Abstract

Early life nutrition has substantial influences on postnatal health, with both under- and overnutrition linked with permanent metabolic changes that alter reproductive and immune function and significantly increase metabolic disease risk in offspring. Since perinatal nutrition depends in part on maternal metabolic condition, maternal diet during gestation and lactation is a risk factor for adult metabolic disease. Such developmental responses may be adaptive, but might also result from constraints on, or pathological changes to, normal physiology. The rising prevalence of both obesity and osteoporosis, and the identification of links among bone, fat, brain, and gut, suggest that obesity and osteoporosis may be related, and moreover that their roots may lie in early life. Here we focus on evidence for how maternal diet during gestation and lactation affects metabolism and skeletal acquisition in humans and in animal models. We consider the effects of overall caloric restriction, and macronutrient imbalances including high fat, high sucrose, and low protein, compared to normal diet. We then discuss potential mechanisms underlying the skeletal responses, including perinatal developmental programming via disruption of the perinatal leptin surge and/or epigenetic changes, to highlight unanswered questions and identify the most critical areas for future research.

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Introduction

Beginning with the observation that individuals born at low birthweight have increased risk of cardiovascular disease [1], there is increasing evidence that early life nutrition influences postnatal health. Both low birthweight [2–10] and high birthweight [11–14] are associated with reduced reproductive and immune function, and significantly higher likelihood of obesity, atherosclerosis, type II diabetes, and the metabolic syndrome in adulthood [15], suggesting that poor intrauterine nutrition might be a risk factor for adult metabolic disease [16,17] (Fig. 1). For example, children of women who were pregnant during the Dutch Hunger Winter of 1944–1945 have increased incidence of obesity, hyperlipidemia, and atherosclerosis if their mothers were impacted by famine during the first trimester; higher rates of pulmonary and kidney disease if during the second trimester; and impaired glucose tolerance if during the third trimester of pregnancy [18–20]. In comparison, children of obese
The relationship between perinatal nutrition and postnatal metabolic disease may be U-shaped, with higher risk of metabolic dysregulation in both perinatal under- and overnutrition.

Adapted from Fig. 1 in Grattan, D. R. Endocrinology 2008;149:5345–5347.

mothers, or those born large for gestational age, are also at increased risk of developing the metabolic syndrome compared to children born at normal birthweight from normal weight mothers, particularly if their mothers also had gestational diabetes mellitus [13]. While these associations are striking, they do not explain why in utero nutrition would alter perinatal metabolism, nor reveal the mechanisms involved. The Barker Hypothesis [21], now known as the Developmental Origins of Adult Health and Disease model, posits that the perinatal environment induces lasting changes in physiological parameters such as hormone levels, glucose tolerance, and satiety, a phenomenon known as developmental programming. The hypothesis is that such ‘programming’ allows offspring to use maternal cues to adapt to the likely postnatal nutritional environment. For example, women born at lower birthweight have lower estrogen levels and are more susceptible to ovarian suppression during energetic stress compared to women born at higher birthweight [22], suggesting restricted perinatal energy availability results in increased sensitivity to adult energetic stress. However, when perinatal and postnatal environments are mismatched, offspring are at increased risk of metabolic disease, a model known as the Predictive Adaptive Response Hypothesis [9,23]. For example, Hales and Barker demonstrated a strong association of low birthweight and type II diabetes risk [4], and proposed that this phenomenon resulted from offspring developing a “thrifty” phenotype in expectation of food scarcity [24]. Similarly, there appears to be a greater risk of obesity in children with low birthweight who exhibit rapid postnatal catch-up growth, compared to children who grow more slowly [25,26], although these relationships are complex [27]. Alternative models include the Maternal Capital Hypothesis, which suggests that offspring exhibit plasticity in early growth in order to match their energetic needs to maternal metabolic resources, and the Intergenerational Phenotypic Inertia Hypothesis, which posits that offspring adapt not to the transient intrauterine nutritional environment, but rather to the mother’s long-term energetic history [28–30].

Although there is some support for each of these hypotheses, the idea that the metabolic changes induced by perinatal developmental programming are adaptive is itself a hypothesis. As Ellison and Jasienska [31] point out, such developmental responses may be adaptations, but might also result from constraints on, or pathological changes to, normal physiology. Thus the notion that a given trait is an adaptation is a hypothesis that must be tested against the alternative hypotheses that the trait arose through pathology or constraint. The most rigorous approach for discriminating among these alternatives is to develop falsifiable predictions for each hypothesis – adaptation, pathology, or constraint – that can be evaluated against the data [31]. As an example, we might expect different patterns of changes in offspring body size, bone mass, and body composition in response to maternal caloric restriction depending on whether this response is an adaptation to expected postnatal energy restriction, the result of a constraint on intrauterine energy that is released after birth, or the product of permanent pathological disruption of growth processes (Table 1).

Whether the changes are adaptive or not, it is clear that early life nutrition alters postnatal metabolism, particularly adiposity and glucose tolerance. Recent interest has focused on the possibility that the perinatal environment also affects skeletal health [32]. The rising prevalence of both obesity and osteoporosis, and the identification of common mechanisms linking skeletal and metabolic homeostasis, suggest that obesity and osteoporosis may be related disorders, and moreover that their roots may lie in early life (Fig. 2). Recent studies in mice demonstrate that interconnections of bone and fat, as well as brain and gut, play a major role in perinatal glucose homeostasis, fat mass and bone mass, raising the possibility that these mechanisms also affect human metabolism [33,34]. Osteoblasts and adipocytes derive from the same population of mesenchymal stem cells (MSCs), such that increased commitment of MSCs to the adipocytic over the osteoblastic lineage might shift the balance between fat mass and bone mass [35–37]. Osteoblast-derived uncarboxylated osteocalcin increases insulin sensitivity and reduces fat mass in mice; the adipokine leptin suppresses this effect by favoring carboxylation of osteoblast-derived osteocalcin, decreasing insulin sensitivity and insulin secretion [38,39]. Leptin also reduces murine bone mass by inhibiting the anabolic effects of brain-derived serotonin, and by increasing sympathetic tone via beta-adrenergic receptors on osteoblasts [40–42]. Fat-derived peroxisome proliferator-activated receptor (Ppar)-gamma increases marrow fat and decreases bone mass [43], while the protein LDL-receptor related protein 5 (LRPS), which plays a crucial role in bone remodeling, increases bone mass by suppressing production of gut-derived serotonin in rodent models [44]. Thus an increase in adiposity has the potential to decrease bone mass, at least in animal models.

How might maternal diet and body composition affect these interactions between offspring bone and fat mass? It is clear that during postnatal life, specific micronutrients, such as Vitamin D and calcium, are essential for proper skeletal development and bone mass acquisition [45,46]. However, the perinatal influences of these micronutrients on skeletal health are more difficult to establish. In addition to micronutrient intake, mothers may be overnourished or undernourished before Table 1

<table>
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<tr>
<th>Perinatal exposure:</th>
<th>Caloric restriction</th>
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<td>Postnatal response:</td>
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<td>Body size</td>
<td>Smaller than normal, in anticipation of low food availability</td>
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<td>Bone mass</td>
<td>Lower than normal, but appropriate for smaller body size</td>
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<td>% Body fat</td>
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pregnancy, and may overeat or undereat relative to their metabolic needs during gestation and lactation. Even when overall caloric intake is appropriate, the proportion of macronutrients – carbohydrate, protein, and fat – may not be optimal, and maternal diets high in fat or sucrose, or low in protein, have all been associated with offspring metabolic abnormalities [47–49]. However, less is known about the effects of dietary macronutrient profile or overall caloric availability (sufficient or insufficient) on offspring bone mass. The timing of exposure to diet also appears to be critical. For example, caloric restriction in adults may be beneficial to skeletal health, while perinatal or early postnatal caloric restriction is deleterious. Here we focus on evidence for how maternal diet during gestation and lactation affects metabolism and skeletal acquisition in humans and in animal models. We consider the effects of overall caloric restriction, macronutrient imbalances including high fat, high sucrose/maternal diabetes, and low protein, and low Vitamin D, compared to normal diet. We then consider potential mechanisms to highlight unanswered questions and identify the most critical areas for future research.

Maternal body composition, diet, and offspring bone mass

Several studies support the hypothesis that the perinatal environment, as reflected by birthweight, influences postnatal bone mass and/or bone size in humans. For example, in young adults, birthweight is positively associated with bone size and bone mineral content, after controlling for current height, weight, and age [50,51]. In the Hertfordshire cohort study, birthweight is positively associated with adult spine and hip BMC, although the effect is modest, explaining only 1–4% of variation in the adult bone properties [52]. Femoral growth velocity in late gestation, presumably related to maternal nutrition, explains about 10% of the variation in the whole body bone area (excluding the head) at 4 yrs of age in the Southampton Women's Study cohort [53]. However, association does not prove causation. Birthweight depends in part on maternal diet and energy availability during gestation, but this association may be confounded by genetics, placental function, smoking, and socioeconomic status. Thus it is instructive to consider twin studies, in which genetic and environmental factors are likely similar within pairs. For example, in female monozygotic twins, Antoniades et al. [54] report a significant association of birthweight with adult BMD at weight-bearing sites (spine and hip), but not the non-weight bearing forearm.

Undernutrition

Fetal undernutrition, resulting from maternal caloric restriction or from placental abnormalities that restrict fetal blood supply, is linked to decreased bone mass. Low maternal fat stores and vigorous maternal exercise are also associated with lower neonatal bone mass in humans [5], and infants born small for gestational age (SGA) have lower bone turnover markers vs. larger infants [55]. These differences persist into adulthood, as individuals who were born at very low birthweight have lower BMD at the forearm [56] and at the lumbar spine and femoral neck [57] compared to adults born at higher birthweights. However, as with the general association between birthweight and adult bone mass described above, the effect of low birthweight on bone mass may reflect postnatal conditions and/or reduced adult body mass. For example, adults born prematurely have lower BMD vs. adults born at full term, but their BMD is appropriate for their shorter stature [58]. Similarly, low birthweight is associated with early puberty in humans [59], which may decrease adult size and bone mass by prematurely ending skeletal growth.

In animal models, whereas it is clear that maternal calorie restriction or fetal growth restriction increases the likelihood of postnatal hyperphagia, obesity, and insulin resistance [49], the skeletal effects are less well established. Maternal calorie restriction in C57Bl/6j and A/J mice leads to reduced whole body BMC in B6 but not A/J offspring at 6 months of age [60], suggesting strain differences in sensitivity to maternal diet. Rats exposed to intrauterine growth restriction via uterine artery ligation are shorter, with narrower bones, lower whole body and femoral BMC, and lower bone strength in sensitivity to maternal diet. Rats exposed to intrauterine growth restriction via uterine artery ligation are shorter, with narrower bones, lower whole body and femoral BMC, and lower bone strength. For example, in mice, calorie restriction impairs skeletal acquisition if initiated at weaning, but causes fewer skeletal deficits and may actually reduce age-related bone loss if initiated in adulthood [63–65]. In addition,
more work is needed to define optimal approaches to maximize the
catch-up in skeletal acquisition following early life caloric restriction.
In humans with stunted childhood growth [66] or anorexia nervosa
[67], adult skeletal deficits remain despite refeeding, suggesting that
energetic deficits during critical windows of skeletal ontogeny may
not be reversible.

Overnutrition

In humans, fetal overnutrition due to maternal high fat diet and/or
preexisting obesity, or from maternal preexisting or gestational
diabetes, is also associated with skeletal changes, although the effect
of large size for gestational age (LGA) on bone properties may depend
on the root cause, and on whether overnutrition continues postna-
tally. Human LGA infants generally have higher total body BMC and
BMD vs. non-LGA infants, even relative to their higher body mass [68–
70]. In contrast, offspring of diabetic mothers, who are frequently born
LGA, have reduced total body BMD vs. controls [71], although bone
turnover markers in amniotic fluid, fetal and maternal blood are
normal [55,72]. From 5–18 yrs of age, children of mothers with type 1
diabetes tend to be both taller and heavier vs. controls, with higher
total body fat mass; however, both BMC and total body bone area are
higher, such that BMD does not differ from controls [73]. Thus
although high maternal glucose appears to increase offspring size at
birth and continuing into childhood, fetal bone turnover is normal,
and childhood bone mass appears appropriate for body mass. In
comparison, LGA resulting from maternal obesity or high fat diet may
be more deleterious. There is a negative association between maternal
fat mass and offspring BMD in childhood [74], and between maternal
fat intake in the 3rd trimester and offspring femoral neck and lumbar
spine BMD at age 16 [75]. Further, mothers consuming a “healthy”
diet in pregnancy (low intake of sugar, fat, and processed foods) have
offspring with up to 6% greater BMD at 9 yrs of age, compared to
mothers consuming more processed foods [76].

In animal models, maternal diabetes in rats is associated with reduced
calcium content and delayed skeletal mineralization vs. controls [77].
Maternal high fat diet is frequently used to induce metabolic syndrome in
rodent offspring [78,79], although there are conflicting data on whether
maternal obesity must be present prior to pregnancy in order to induce
a metabolic phenotype [80], or whether its effects are similar whether
the diet is long term or confined to pregnancy and lactation [47]. In
mice, offspring of dams exposed to high fat diet are shorter, with lower
total body bone mass at fetal day 19 [81]. Offspring of mothers fed a high
fat diet that are weaned onto the same high fat diet have higher
marrow adiposity, greater body mass, and shorter femurs with larger
cross-sectional dimensions at 30 wks of age [82]. However, it is unclear
whether these differences would remain after adjustment for their
higher body weight. Altogether, whereas studies in animals consistently
show that maternal high fat diet leads to increased risk of metabolic
disturbances in offspring, the effects on skeletal health are less clear.

Micronutrient intake

Maternal micronutrient intake may also impact offspring bone
health, although the data are difficult to interpret for several reasons.
First, the definition of “adequate” maternal vitamin D levels during
pregnancy remains contentious [83]. Second, while some studies
report an association of infant vitamin D levels and BMC [84,85],
others do not, even in extreme vitamin D deficiency. For example, in
the absence of vitamin D supplements, total body BMC is higher in
summer-born than in winter-born Korean infants [86]; in the US,
where vitamin D supplementation is more common, BMC is actually
higher in winter-born vs. summer-born infants [87–89]. On the other
hand, infants with 1-alpha-hydroxylase deficiency or vitamin D
receptor mutations, who cannot synthesize or bind vitamin D, have
normal skeletal phenotypes at birth and can be treated with calcium,
bypassing vitamin D entirely and suggesting it is not essential for
normal bone mineralization [90]. Thus, although it has been reported
that offspring of mothers with 25-hydroxyvitamin D levels < 25 nM/L
during pregnancy have lower total body and lumbar spine BMC at 9
years of age, compared to offspring of mothers with vitamin D levels
>50 nM/L during pregnancy [91], it is not clear that this association
involves developmental programming.

The associations between postnatal bone health and intrauterine
exposure to other micronutrients are similarly modest. For calcium, a
study in the Gambia reported no difference in whole body BMC in the
first year of life in infants of supplemented vs. non-supplemented
women [92]. However, in Indian women, maternal dietary calcium
intake in pregnancy is associated with higher offspring BMD at 6 years
of age [93]. Finally, in the Avon Longitudinal Study of Parents and
Children (ALSPAC) cohort, maternal magnesium, potassium, and
folate consumption in pregnancy are positively associated with bone
properties at 9 years of age, although only the latter remains
significant after body size adjustment [94].

To summarize, these studies demonstrate that perturbations of
maternal caloric intake and/or dietary macronutrient composition in
gestation and lactation have lasting effects on offspring skeletal
acquisition and maintenance, particularly in animal models. However,
the extent to which these changes occur via perinatal developmental
programming or via discordant perinatal vs. postnatal somatic growth
remains to be determined, particularly in humans. Nonetheless, when
the human and animal data are taken together, what is striking is that
any imbalance in maternal diet – too few calories, too much fat, too
little protein – seems to alter postnatal bone mass and microarchi-
tecture. Perinatal exposure to calorie restriction or low protein diet is
deleterious to both cortical and trabecular bone, while exposure to
high fat diet appears to have complex positive and negative effects on
different aspects of trabecular and cortical microarchitecture, though
more data are needed.

Mechanisms that may underlie the skeletal response to altered
perinatal nutrition

The findings reviewed above demonstrate the need for additional
studies in humans to test the hypothesis that the perinatal
environment sensitizes the fetus to the expected postnatal nutritional
environment [95], which, for the skeleton, might involve a reduction
in bone mass that decreases energy requirements but increases
osteoporosis risk [96]. The question is whether the mechanisms
involve direct perinatal developmental programming of bone, as has
been shown for metabolic phenotypes, and/or involve other mech-
isms or secondary effects of perinatal programming of other tissues
that then influence skeletal metabolism. Given the limited number of
studies addressing this question, particularly for bone, any effort to
identify mechanisms is necessarily speculative and relies heavily on
data from animal studies. Furthermore, there are many potential
factors that could underlie the effects of perinatal diet on skeletal
acquisition, including changes in the growth hormone/insulin-like
growth factor (IGF-1) axis [97,98], or alterations in glucocorticoid
levels, such as cortisol [99]. Two other strong candidate hormonal
mediators of bone-diet interactions are the adipokine leptin, which
may affect bone mass directly [100–102], via hypothalamic-mediated
ß-adrenergic signaling [42,103,104], or via suppression of brain-
derived serotonin production [41]; and osteocalcin, which has
recently been identified as a bone-derived hormone that controls
energy metabolism [105,106]. Some data demonstrate that osteocal-
cin is produced by adipocytes as well as by osteoblasts, suggesting
both fat and bone may contribute to glucose homeostasis via
osteocalcin production [107]. There may also be direct effects of
nutrition on bone cell number, proliferation, and growth rate
[32,168,109]. Here we focus on two potential mechanisms that are
currently under investigation: perinatal developmental programming
of postnatal leptin levels, and epigenetic changes due to genomic imprinting or DNA methylation.

Perinatal developmental programming of hormone levels

Although the mechanisms involved in developmental programming of metabolism are incompletely understood, one hypothesis is that maternal undernutrition or overnutrition disrupts the perinatal leptin surge, altering hypothalamic development and postnatal regulation of food intake and metabolism [110,111]. In rodents, the perinatal leptin surge promotes neuronal connections between the arcuate and paraventricular nuclei, particularly neuropeptide Y (Npy) and agouti-related protein (AgRP) projections [112,113], and sensitizes hypothalamic neurons to postnatal leptin [114–116]. Maternal caloric restriction in rodents triggers this leptin surge prematurely, resulting in postnatal obesity, peripheral leptin resistance, and impaired leptin transport to the brain [117–119]. Similarly, delay and inhibition of the perinatal leptin surge in a rat model of intrauterine growth restriction causes hypothalamic disorganization [120]. Neonatal leptin treatment rescues postnatal metabolism in mouse pups exposed to maternal calorie restriction, but causes obesity and metabolic problems despite isocaloric intake in offspring exposed to maternal normal diet [121], whereas leptin antagonist treatment in early postnatal life mimics the effects of maternal caloric restriction, causing hyperleptinemia and obesity [122]. Similarly, maternal caloric restriction triggers insulin resistance and obesity in postnatally ad libitum-fed mice, but not in leptin-treated or postnatally calorie-restricted mice [49,114].

In maternal overnutrition, the mechanisms involved in developmental programming are less clear. In rodent models, it has been suggested that neonatal hyperleptinemia alters leptin receptor expression and contributes to selective leptin resistance in the arcuate nucleus of the hypothalamus, as is seen in postnatal hyperleptinemia [15]. High fat diet has also been shown to promote the growth of neurons expressing orexigenic peptides in rats [123], which could increase fat mass and consequently leptin levels. Increased fat intake by rat dams during late pregnancy and lactation is associated with higher offspring fat mass and leptin levels at weaning, but lower leptin levels than controls thereafter [124].

Unfortunately, the existence and timing of a perinatal leptin surge in humans remain unclear, although it appears that fetal leptin levels are high just prior to birth and fall in the first week of life. What is clear is that perinatal undernutrition or overnutrition alters postnatal leptin levels. Infants who are born large for gestational age (LGA) have higher than normal leptin levels [125,126], which persist into childhood [127]. Conversely, children born small for gestational age (SGA) have abnormally low leptin levels at birth [128,129] and in childhood [130]. Further, breast milk leptin levels vary with maternal diet, and it has even been suggested that this variation – and the absence of leptin in infant formula – may contribute to the protective effect of breastfeeding on postnatal obesity [131,132]. However, the possible ramifications of leptin levels on skeletal acquisition are unclear. While some studies report a positive association between neonatal skeletal size, bone mineral density and umbilical cord blood leptin levels [133], others find no such association [69].

How might programming of leptin circuits in the brain affect postnatal bone mass? Leptin has complex effects on trabecular and cortical bone, both directly and via the hypothalamus. In wildtype mice, intracerebroventricular (iCV) leptin infusion increases sympathetic tone via binding to hypothalamic receptors, which then activates β-adrenergic receptors in osteoblasts, stimulating bone resorption and decreasing bone formation [101,103,104]. However, a recent study reports that iCV leptin increases bone formation in the leptin-deficient ob/ob mouse [134]. In addition, there is some evidence that leptin has peripheral anabolic effects, promoting commitment of human-derived mesenchymal stem cell lines (MSCs) to bone vs. fat lineages, and increasing periosteal bone formation and inhibiting ovariectomy-induced bone loss in rodent models [100,102,135]. However, a subsequent study in mice found that leptin receptor (Lepr) deletion in osteoblasts did not alter skeletal phenotype, suggesting leptin has no direct effect on osteoblasts [136]. Accordingly, there are at least two mechanisms by which perinatal programming could affect postnatal skeletal development by altered leptin levels: 1) via direct effects on osteoblast progenitors and mature osteoblasts, and/or 2) via alterations in leptin-induced, hypothalamic-mediated β-adrenergic signaling in bone. In addition to these potential direct mechanisms, there are likely indirect effects of leptin on bone homeostasis via modulation of other central or peripheral hormones, such as osteocalcin and adiponectin [137]. However, it is not clear whether more leptin is always better for bone. Injections of leptin in pregnant rats, which should signal greater energy availability, actually reduce offspring postnatal skeletal growth and cortical bone mass, as well as adiposity [138]. Furthermore, one recent study has challenged the notion that perinatal leptin is primarily responsible for postnatal metabolic dysregulation. In ob/ob mice, which lack leptin, pups exposed to maternal calorie restriction followed by catch-up growth have higher body mass than controls, demonstrating that programming induced by maternal diet does not occur through leptin alone [139].

While the concept of perinatal developmental programming of bone is an intriguing one, more data are needed to establish whether the skeletal changes induced by maternal diet involve programming of leptin or other hormones, and to understand how interactions of leptin levels in the perinatal, lactational, and post-weaning time frames affect postnatal skeletal acquisition and maintenance.

Epigenetic changes

Epigenetic changes in gene expression, such as altered DNA methylation, have been implicated in a wide range of human diseases, from cancer [140] to allergy and asthma [141] to the function of specific genes including the glucocorticoid receptor gene [142]. Given the enormous potential of this mechanism to explain how perinatal influences might cause lifelong changes in offspring physiology, interest in potential epigenetic effects of maternal diet on offspring bone mass is high. Although this research is in its early stages, there are some tantalizing hints. For example, perinatal famine exposure, as in the Dutch Hunger Winter, has been associated with altered methylation of the IGF2 gene in adulthood [143], which could influence bone mass.

Diet has also been shown to alter postnatal transcriptional activity of multiple genes that may affect bone mass. In rats, offspring of mothers exposed to high fat diet have altered postnatal expression of the leptin receptor (ObRb), pro-opiomelanocortin (Pomc), neuropeptide Y (Npy), and signal transducer and activator of transcription 3 (Stat3) [144,145]. Perinatal calorie restriction combined with postnatal high fat diet in rats is also associated with significant changes in Pomc, AgRP, Npy, and ObRb expression, along with high leptin and insulin levels [146]. Offspring of mothers exposed to high fat diet also have increased preference for sugary and fatty foods, and show DNA hypomethylation in genes associated with reward, including opioids and dopamine [147]. At the other extreme of energy availability, offspring from calorie-restricted rats have altered expression of genes involved in lipid metabolism and glucose homeostasis, including higher expression of 11β-hydroxysteroid dehydrogenase type 2 (11β-HSD2), lower expression of peroxisome proliferator-activated receptor (Ppar)-alpha, the glucocorticoid receptor, and phosphoenolpyruvate carboxykinase (Pepck), and exhibit increased methylation of the PPARα promoter and glucocorticoid receptor genes, but neonatal leptin treatment normalizes both gene expression and metabolic phenotype [148].

Among the most intriguing observations is that high fat diet-induced changes can persist for multiple generations via epigenetic mechanisms. Four generations of mice continuously raised on high fat diet exhibit progressively increasing fat mass despite no changes in food intake, along with changes in gene expression including colony-
stimulating factor 3 (also known as granulocyte colony-stimulating factor) and Nocturnin [149]. Both male and female mice exposed to maternal high fat and/or overfeeding during lactation can transmit reduced insulin sensitivity and increased body length to their offspring through epigenetic inheritance [150,151]. These studies provide a template for understanding how perinatal diet might influence offspring metabolism, and illustrate the importance of epigenetic mechanisms in moderating between the individual and the environment. The great challenges going forward will be to determine whether epigenetic regulation has direct effects on offspring bone mass and microarchitecture, and whether deleterious epigenetic changes might be modified postnatally to reduce disease risk.

Unanswered questions and future directions

Due to the growing evidence that perinatal programming influences metabolic and cardiovascular disease risk in humans, there is tremendous interest in understanding how the perinatal environment, including maternal diet and other factors such as smoking, exercise, and body mass, influence postnatal skeletal acquisition and maintenance. Studies in animal models demonstrate that both undernutrition and overnutrition, as well as low protein intake, are deleterious to offspring bone mass. In humans, birthweight, as a surrogate for maternal perinatal nutrition, is linked to adult bone mass, but the proportion of variance in bone mass explained by birthweight tends to be small. It is also unclear whether this association results from perinatal developmental programming of postnatal skeletal acquisition, or simply reflects the normal relationship between skeletal size and bone mass. However, while perinatal developmental programming of the skeleton in humans remains to be definitively established, there is some evidence for an influence of developmental programming of bone in animal models. Further studies are needed to determine whether the effects of perinatal programming on the skeleton are direct effects on bone cells and/or bone metabolism, or are indirect effects due to alterations in metabolism and energy utilization.

Maternal perinatal diet may influence offspring bone mass via perinatal developmental programming of leptin levels, epigenetic mechanisms such as DNA methylation or other types of genomic imprinting, or other mechanisms such as direct effects on bone cell differentiation, proliferation or gene expression. More data on the nature of a perinatal leptin surge in humans, as well as a better understanding of how the perinatal leptin surge influences bone mass, would be particularly helpful. Another key area for future research is the observation that the effects of maternal diet vary in gestation versus lactation, particularly in terms of neuronal development and programming of feeding circuits. For example, maternal calorie restriction in rats during gestation only, followed by rapid catch-up growth in lactation, is associated with obesity and hyperleptinemia, while offspring of mothers whose calorie restriction continued through lactation do not exhibit these abnormalities [152]. Understanding the relative importance of diet during gestation vs. lactation in establishing postnatal bone mass may shed light on the mechanisms involved.

Finally, the hypothesis that perinatal developmental programming of skeletal phenotypes is adaptive should be evaluated in a rigorous hypothesis testing framework that considers pathology and constraint as alternatives to adaptation [31]. Once the mechanisms underlying these developmental responses have been established, the next goal will be to develop strategies to improve bone mass and reduce osteoporosis risk in affected individuals.

References


